



CGCTCAGGATACGACTTCCTGCTAGAGGATCGGATCCCGGGGCTGATTATATAGCTCGATCGATC
 TTCTCTATATCTGGGCTGGGCTATATACACACACACCTCGCGGATAGCATGACTGATCTA
 CCCCCT
 CACAGACTTACGGCT

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Features

☐ 1: Z70522. C.melo mRNA (clon...[gi:1843441]

Links

LOCUS CMPMEL7 686 bp mRNA linear PLN 22-JUN-2001
 DEFINITION C.melo mRNA (clone pMel7).
 ACCESSION Z70522
 VERSION Z70522.1 GI:1843441
 KEYWORDS AEl gene.
 SOURCE Cucumis melo (muskmelon)
 ORGANISM Cucumis melo
 Eukaryota; Viridiplantae; Streptophyta; Embryophyta; Tracheophyta;
 Spermatophyta; Magnoliophyta; eudicotyledons; core eudicots;
 rosids; eurosids I; Cucurbitales; Cucurbitaceae; Cucumis.
 REFERENCE 1 (bases 1 to 686)
 AUTHORS Aggelis,A., John,I., Karvouni,Z. and Grierson,D.
 TITLE Characterization of two cDNA clones for mRNAs expressed during
 ripening of melon (Cucumis melo L.) fruits
 JOURNAL Plant Mol. Biol. 33 (2), 313-322 (1997)
 MEDLINE 97188564
 PUBMED 9037149
 REFERENCE 2 (bases 1 to 686)
 AUTHORS Aggelis,A., John,I., Karvouni,Z. and Grierson,D.
 TITLE Characterization of two cDNA clones for mRNAs expressed during
 ripening of Melon (Cucumis melo L.) fruits
 JOURNAL Unpublished
 REFERENCE 3 (bases 1 to 686)
 AUTHORS John,I.
 TITLE Direct Submission
 JOURNAL Submitted (01-APR-1996) John I., The University of Nottingham,
 Physiology and Environmental Science, Sutton Bonington Campus,
 Loughborough, United Kingdom, LE12 5RD

FEATURES

Location/Qualifiers

source

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 1..30
 31..486
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 /db_xref="GI:1843442"
 /db_xref="SPTREMBL:P93095"

mRNA

5' UTR

CDS

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AYQFVPKDRNHCQAILSIEYEKLHHGSPDPHKYIDLMITNDIGSHIK"

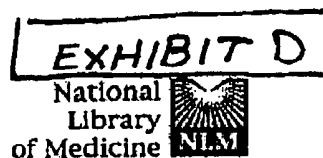
3'UTR 487..686
polyA_signal 646..651
polyA_site 686

BASE COUNT 211 a 97 c 135 g 243 t
ORIGIN

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121 gttcccaata taacccccag atgcattcaa caagttgaaa ttcattggtac taattgggat
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301 ggagatgtgt tcaaaaatta taaaagcttt aaaccagctt accaatttgt acctaaggat
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661 tcatcccttg tttatgtttc gttatt

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1: Nucleic Acids Res. 1991 Apr 11;19(7):1571-6.

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Upstream sequences of rice proliferating cell nuclear antigen (PCNA) gene mediate expression of PCNA-GUS chimeric gene in meristems of transgenic tobacco plants.

Kosugi S, Suzuka I, Ohashi Y, Murakami T, Arai Y.

Institute of Applied Biochemistry, University of Tsukuba, Ibaraki, Japan.

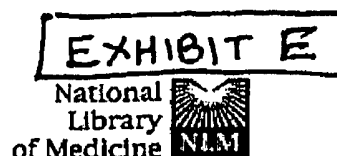
The transgenic tobacco plants have been generated that express the E. coli beta-glucuronidase (GUS) gene under control of the promoter from the rice proliferating cell nuclear antigen (PCNA, DNA polymerase auxiliary protein gene. GUS expression detected in situ by staining with the chromogenic substrate, 5-bromo-4-chloro-3-indolyl-beta-D-glucuronide (X-Gluc), was restricted to meristems in the organs of the transgenic tobacco plants. This expression responded to the phytohormones which promote callus formation. Furthermore, in situ thymidine uptake showed that the GUS expression pattern corresponded well to the active sites of DNA synthesis. Deletion analysis of the 5' upstream sequence confined the GUS expression pattern to fragment extending 263 bp upstream of the transcription start site of the rice PCNA gene. Thus, we have identified this fragment as a main regulatory element of the rice PCNA gene promoter.

PMID: 1709277 [PubMed - indexed for MEDLINE]

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1: Plant Physiol. 1994 May;105(1):357-67.

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Identification of a light-responsive region of the nuclear gene encoding the B subunit of chloroplast glyceraldehyde 3-phosphate dehydrogenase from Arabidopsis thaliana.

Kwon HB, Park SC, Peng HP, Goodman HM, Dewdney J, Shih MC.

Department of Biological Sciences, University of Iowa, Iowa City 52242.

We report here the identification of a cis-acting region involved in light regulation of the nuclear gene (GapB) encoding the B subunit of chloroplast glyceraldehyde 3-phosphate dehydrogenase from Arabidopsis thaliana. Our results show that a 664-bp GapB promoter fragment is sufficient to confer light induction and organ-specific expression of the Escherichia coli beta-glucuronidase reporter gene (Gus) in transgenic tobacco (Nicotiana tabacum) plants. Deletion analysis indicates that the -261 to -173 upstream region of the GapB gene is essential for light induction. This region contains four direct repeats with the consensus sequence 5'-ATGAA(A/G)A-3' (Gap boxes). Deletion of all four repeats abolishes light induction completely. In addition, we have linked a 109-bp (-263 to -152) GapB upstream fragment containing the four direct repeats in two orientations to the -92 to +6 upstream sequence of the cauliflower mosaic virus 35S basal promoter. The resulting chimeric promoters are able to confer light induction and to enhance leaf-specific expression of the Gus reporter gene in transgenic tobacco plants. Based on these results we conclude that Gap boxes are essential for light regulation and organ-specific expression of the GapB gene in A. thaliana. Using gel mobility shift assays we have also identified a nuclear factor from tobacco that interacts with GapA and GapB DNA fragments containing these Gap boxes. Competition assays indicate that Gap boxes are the binding sites for this factor. Although this binding activity is present in nuclear extracts from leaves and roots of light-grown or dark-treated tobacco plants, the activity is less abundant in nuclear extracts prepared from leaves of dark-treated plants or from roots of greenhouse-grown plants. In addition, our data show that this binding factor is distinct from the GT-1 factor, which binds to Box II and Box III within the light-responsive element of the RbcS-3A gene of pea.

PMID: 8029358 [PubMed - indexed for MEDLINE]